

EXPLORING INHIBITORY EFFECTS OF *Crataevanurvala*-DERIVED SILVER NANOPARTICLES ON HIGH RESISTANCE BACTERIAL AND FUNGAL PATHOGENS

Deepak Kumar^{1,✉}, Ashwani Sanghi², Versha Parcha¹, Shefali Arora³,
Mohd. Danish¹, Abdullah¹ and Aditya Swarup⁴

¹Department of Pharmaceutical Chemistry and Chemistry, Dolphin (PG) Institute of Biomedical and Natural Sciences, Dehradun, Uttarakhand, India.

²School of Allied Health Sciences, MVN University, Haryana, India.

³Department of Chemistry, University of Petroleum and Energy Studies, Dehradun, Uttarakhand, India

⁴Department of Medical Lab Technology, Dolphin (PG) Institute of Biomedical and Natural Sciences, Dehradun, Uttarakhand, India

✉Corresponding author: deepsingh2304@gmail.com

ABSTRACT

Silver nanoparticles are increasingly being utilized as antibacterial as well as antifungal agents in several applications. The use of antibiotics has emerged microbe resistance among multidrug-resistant organisms, which is endangering the global health community. Plants are a good source of phytoconstituents and have been successfully applied for the treatment of microbial diseases. Thus, silver nanoparticles from the ethanol extract of leaves of *Crataeva nurvala* were biosynthesized and aimed to explore the antibacterial and antifungal effects. The antibacterial and antifungal properties of the ethanol extract and its silver nanoparticles were evaluated at concentrations of 200 mg/ml and 50 mg/ml, respectively, using the well diffusion technique. Ethanol extract and its silver nanoparticles from *Crataeva nurvala* showed antibacterial activity against *E. coli*, *Bacillus cereus*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* and antifungal activity against *Aspergillus niger*, *Rhizopus oryzae*, *Mucor azygosporus* and *Penicillium chrysogenum*. Antibacterial and antifungal effects of silver nanoparticles were significantly higher than compared of ethanol extract and standard drugs. Hence, findings of the research suggested that the silver nanoparticles from *Crataeva nurvala* leaves exhibited antibacterial and antifungal effects, which might help to develop a new drug for treatment against various diseases.

Keywords: *Crataeva nurvala*, Silver Nanoparticles, Antibacterial, Antifungal, Well Diffusion.

RASAYAN J. Chem., Vol. 18, No.3, 2025

INTRODUCTION

The overuse of antibiotics led to microbial resistance worldwide and represents a major threat to humanity.¹ It results in an increased demand for the discovery of novel antibacterial and antifungal agents.² WHO has identified antimicrobial resistance as one of the top ten worldwide public health concerns.³ Nonetheless, plants are frequently utilized in rural regions as a remedy for a variety of illnesses, and because they are abundant in bioactive substances like antimicrobials, which can help to treat difficult bacterial infections.⁴ Since plants contain secondary metabolites with a variety of biological activities, like antibacterial, antioxidant, antifungal, and anti-inflammatory properties, they have long been used as prophylactics.^{5,6} The use of herbal medicine has increased due to the side effects of synthetic drugs, as well as resistance to drugs in a specific treatment.^{7,8} The nanoparticles have gained large interest for the past two decades as their physico-chemical properties, controlled size and shape, and ability to interact with other molecules.⁹ Apart from several applications of nanoparticles, antibacterial and antifungal uses of nanoparticles are expected to be fruitful in treating bacterial and fungal illnesses, and also in multidrug resistance.¹⁰ The biosynthesized nanoparticles can enhance the effect of phytoconstituents in the extract against microbial activity.¹¹ There are various ways to biosynthesize silver nanoparticles, which are simple, fast, non-toxic, effective, and environmentally sustainable. It has broad-spectrum efficacy and low cytotoxicity and has been shown to be very effective against microorganisms. Plant parts are used to

prepare plant extracts, which are utilized for the biosynthesis of Silver nanoparticles (AgNPs) for their application in numerous sectors, especially in Pharmaceuticals.¹² *Crataeva nurvala* Buch. Ham is found in tropical regions of the world, including India. The plant has been used as a traditional remedy for blood purification, fever, respiratory issues, memory loss, and weakened immune systems. Numerous phytoconstituents, including beta-sitosterol, luteolin, kaempferol, rutin, betulinic acid, and catechin, have been identified in the plant. Various plant parts have been documented to provide therapeutic benefits, including anti-inflammatory, wound-healing, cardioprotective, antipyretic, anti-urolithic, anxiolytic, anti-diabetic, and anti-cancer effects.¹³ Therefore, this research has aimed to investigate the antibacterial and antifungal properties of ethanol extract and its silver nanoparticles.

EXPERIMENTAL

Collection of Plant Materials

Crataevanurvala leaves were collected from the area around the FRI, Dehradun, Uttarakhand, India, and verified by Dr. Vidit Tyagi, a botanist with the Department of Botany at DIBNS, Dehradun.

Preparation of Ethanol Extract of Leaves of *Crataeva nurvala*

The leaves were washed with water and dried in the shade. The air-dried leaves (200 g) were crushed and extracted with ethanol. The extract containing ethanol was evaporated till dryness to obtain the residue and concentrated under reduced pressure.¹⁴

Biosynthesis of AgNPs

AgNPs were biosynthesized using an ethanol extract of *Crataeva nurvala*. An aqueous solution containing 1 mM silver nitrate (AgNO₃) was made. 500 mg of ethanol extract was dissolved with a small amount of ethanol and diluted up to 100 mL with water. 10 ml of the silver nitrate solution was mixed with 60 ml ethanol extract solution and stirred for 2 hrs and kept at room temperature to react.¹⁵

Inhibitory Effects of Ethanol Extract and AgNPs on Bacterial and Fungal Pathogens

The inhibitory studies on bacterial and fungal pathogens were done by the well diffusion method as described by Barry A for ethanol extract and AgNPs.¹⁶ The concentration of ethanol extract was 200 mg/ml, and silver nanoparticles were 50 mg/ml. *Escherichia coli*, *Bacillus cereus*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* were used for antibacterial study. *Aspergillus niger*, *Rhizopus oryzae*, *Mucor azygosporus* and *Penicillium chrysogenum* were used for antifungal study.

RESULTS AND DISCUSSION

Earlier, in continuation with the research, we have reported the characterization of biosynthesized silver nanoparticles and their membrane stabilizing and antioxidant activities.¹⁷

Inhibitory Effects of Bacterial Pathogens

Ethanol extract showed an inhibition zone of 16.83±0.33 mm against *E. coli*, 12.62±0.75 mm against *Bacillus cereus*, 11.75±0.38 mm against *Pseudomonas aeruginosa*, and 16.92±0.22 mm against *Staphylococcus aureus*. AgNPs showed a bacterial inhibition zone of 17.73±0.25 mm against *E. coli*, 13.50±0.58 mm against *Bacillus cereus*, 15.33±0.60 mm against *Pseudomonas aeruginosa*, and 26.32±0.44 mm against *Staphylococcus aureus*. The standard drug Chloramphenicol was taken for study for antibacterial activity. Chloramphenicol showed an inhibition zone of 20.67±0.55 mm against *E. coli*, 17.42±0.68 mm against *Bacillus cereus*, 16.58±0.79 mm against *Pseudomonas aeruginosa*, and 27.12±0.39 mm against *Staphylococcus aureus* (Table-1 and Fig.-1).

Table-1: Results of Inhibitory Effects on Bacterial Pathogens

Bacterial Strains	Inhibitory effects on bacterial pathogens(zone of inhibition in mm)		
	Ethanol extract	AgNPs	Standard Drug Chloramphenicol
<i>E. coli</i>	16.83±0.33	17.73±0.25	20.67±0.55
<i>Bacillus cereus</i>	12.62±0.75	13.50±0.58	17.42±0.68
<i>Pseudomonas aeruginosa</i>	11.75±0.38	15.33±0.60	16.58±0.79
<i>Staphylococcus aureus</i>	16.92±0.22	26.32±0.44	27.12±0.39

Results are expressed as mean values ± standard error (n = 3)

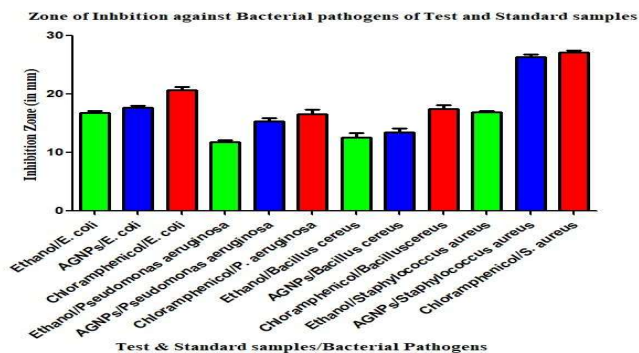


Fig.-1: Results of Zone of Inhibition of Bacterial Pathogens by Ethanol Extract, AgNPs, and Chloramphenicol

Inhibitory Effects of Fungal Pathogens

Ethanol extract showed inhibition zone 15.30 ± 0.39 mm against *Aspergillus niger*, 18.57 ± 0.49 mm against *Rhizopus oryzae*, 11.42 ± 0.57 mm against *Mucor azygosporus* and 9.6 ± 0.43 mm against *Penicillium chrysogenum*. AgNPs showed fungal inhibition zone 17.28 ± 0.21 mm against *Aspergillus niger*, 20.00 ± 0.25 mm against *Rhizopus oryzae*, 19.23 ± 0.52 mm against *Mucor azygosporus* and 12.30 ± 0.22 mm against *Penicillium chrysogenum*. The standard drug Clotrimazole was taken for the study for antifungal activity. Clotrimazole showed inhibition zone 9.27 ± 0.71 mm against *Aspergillus niger*, 11.07 ± 1.08 mm against *Rhizopus oryzae*, 14.70 ± 0.53 against *Mucor azygosporus*, 13.30 ± 0.74 mm against *Penicillium chrysogenum* (Table-2 and Fig.-2).

Table-2: Results of Inhibitory Effects on Fungal Pathogens

Fungal Strains	Inhibitory effects on fungal pathogens(zone of inhibition in mm)		
	Ethanol extract	AgNPs	Standard Drug Clotrimazole
<i>Aspergillus niger</i>	15.30 ± 0.39	17.28 ± 0.21	9.27 ± 0.71
<i>Rhizopus oryzae</i>	18.57 ± 0.49	20.00 ± 0.25	11.07 ± 1.08
<i>Mucor azygosporus</i>	11.42 ± 0.57	19.23 ± 0.52	14.70 ± 0.53
<i>Penicillium chrysogenum</i>	9.6 ± 0.43	12.30 ± 0.22	13.30 ± 0.74

Results are expressed as mean values $\pm$ standard error (n = 3)

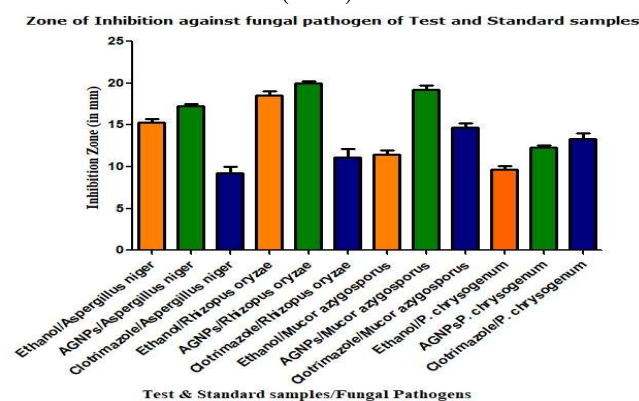


Fig.-2: Results of Zone of Inhibition of Fungal Pathogens by Ethanol Extract, AgNPs, and Clotrimazole

Two very different modes of action should be taken into account when creating antimicrobial materials: first, those that involve direct interactions with bacterial cells, and second, those that facilitate nanomaterial entry into infected host cells to obtain access to intracellular pathogens.¹⁸ While the antimicrobial effect of silver is widely recognized.¹⁹ Regarding AgNPs, the exact mechanism by which nanocrystalline silver causes bacterial cell death is yet unknown, although it probably involves a variety of different ways, mostly due to the release of silver ions (Ag^+).^{20,21} These include changes to the bacterial cell wall and cell membrane, interactions with DNA, binding or inhibition of membrane proteins and enzymes, and elevated reactive oxygen species (ROS) levels.^{21,22} The ability of AgNPs to penetrate

the bacterial cell wall, binding to the cell membrane, changing its permeability and structure, while also modifying transport activity. Furthermore, AgNPs produce silver ions (Ag⁺), which further damage bacterial cell membranes, accumulating inside the cell, and disrupt a number of essential processes.^{20,22}

CONCLUSION

We used an ethanol extract of the leaves of the *Crataeva nurvala* plant to prepare a one-step biosynthesis for AgNPs. The process of biosynthesis is quick, easy, repeatable, and safe for the environment. AgNPs and ethanol extract exhibited antibacterial and antifungal activities. These biosynthesized AGNPs can be used in a variety of pharmaceutical products, including coatings, tablets, creams, ointments, gels, and more.

ACKNOWLEDGMENTS

The authors are thankful to the Management of DIBNS, Dehradun, for providing the necessary facilities for the completion of this work.

CONFLICT OF INTERESTS

The authors declare that there is no conflict of interest.

AUTHOR CONTRIBUTIONS

The present work is related to *SDG-3: Good Health and Well-being*. All the authors contributed significantly to this manuscript, participated in reviewing/editing and approved the final draft for publication. The research profile of the authors can be verified from their ORCID ids, given below:

Deepak Kumar  <http://orcid.org/0000-0002-5608-3883>

Ashwani Sanghi  <http://orcid.org/0009-0006-2619-1433>

Versha Parcha  <http://orcid.org/0000-0003-0202-1625>

Shefali Arora  <http://orcid.org/0000-0002-0312-4127>

Mohd. Danish  <http://orcid.org/0009-0004-9942-4643>

Abdullah  <http://orcid.org/0009-0006-1190-8817>

Aditya Swarup  <http://orcid.org/0009-0006-7754-4470>

Open Access: This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

REFERENCES

- Osama Y. Althunibat, Haitham Qaralleh, Sati Yassin Ahmed Al-Dalin, Muayad Abboud, Khaled Khleifat, Ibrahim S. Majali, Hammad K. H. Aldal'in, Walid A. Rayyan and Ahmad Jaafraa, *Journal of Pure and Applied Microbiology*, **10(1)**, 367(2016).
- Khaled A. Tarawneh, Nafe' M. Al-Tawarah, Adel H. Abdel-Ghani, Ahmed M. Al-Majali, Khaled M. Khleifat, *Journal of Basic Microbiology*, **49(3)**, 310(2009), <https://doi.org/10.1002/jobm.200800060>
- <https://www.who.int/docs/default-source/antimicrobial-resistance/amr-factsheet.pdf>
- T.P. Tim Cushnie, Benjamart Cushnie, Andrew J. Lamb, *International Journal of Antimicrobial Agents*, **44(5)**, 377(2014), <https://doi.org/10.1016/j.ijantimicag.2014.06.001>
- Hercules Sakkas and Chrissanthi Papadopoulou, *Journal of Microbiology and Biotechnology*, **27(3)**, 429(2017), <https://doi.org/10.4014/jmb.1608.08024>
- ć'smail Koyuncu, Ataman Gönel, Abdurrahman Akdağ, Mustafa Abdullah Yilmaz, *Cellular and Molecular Biology*, **64(1)**, 75(2018), <https://doi.org/10.14715/cmb/2018.64.1.14>
- Lakshmi Mohan, *Plant-Based Drugs as an Adjuvant to Cancer Chemotherapy, Alternative Medicine*, Intechopen, UK, (2020), <https://doi.org/10.5772/intechopen.94040>
- Cyrill L. Gorlenko, Herman Yu. Kiselev, Elena V. Budanova, Andrey A. Zamyatnin, Jr. and Larisa N. Ikryannikova, *Antibiotics*, **9(4)**, 170(2020), <https://doi.org/10.3390/antibiotics9040170>
- Krishna Suresh Babu Naidu, Nelisha Murugan, J. K. Adam and Ser Shen, *BioNanoScience*, **9(2)**, 266(2019), <https://doi.org/10.1007/s12668-019-00612-4>

10. Hengyi Xu, Feng Qu, Hong Xu, Weihua Lai, Y. Andrew Wang, Zoraida P. Aguilar and Hua Wei, *BioMetals*, **25(1)**, 45(2012), <https://doi.org/10.1007/s10534-011-9482-x>
11. Anti Islam, Chanchal Mandal, Ahsan Habib, *Journal of Advanced Biotechnology and Experimental Therapeutics*, **4(1)**, 67(2021), <https://doi.org/10.5455/jabet.2021.d108>
12. S. Thota S., D.C. Crans, *Metal nanoparticles: synthesis and applications in pharmaceutical sciences*, John Wiley and Sons, Hoboken, US, 193(2018).
13. D. Kumar, S. Sharma and S. Kumar, *Future Journal of Pharmaceutical Sciences*, **6**, 113(2020), <https://doi.org/10.1186/s43094-020-00106-1>
14. P. Tiwari, B. Kumar, M. Kaur, G. Kaur, H. Kaur, *Internationale Pharmaceutica scientia*, **1(98)**, 106(2011).
15. V. Parashar, R. Parashar, B. Sharma, A. C. Pandey, *Digest Journal of Nanomaterials and Biostructures*, **4(1)**, 45(2009).
16. Arthur L. Barry, Lea. and Fibiger, The antimicrobial susceptibility test principle and practices, *Annals of Internal Medicine*, **87(3)**, 388(1977), https://doi.org/10.7326/0003-4819-87-3-388_1
17. Deepak Kumar. Versha Parcha. Ashwani Sanghi. Mohd. Danish. Abdullah. Aditya Swarup. Shefali Arora, *Rasayan Journal of Chemistry*, **18(1)**, 406(2025), <http://doi.org/10.31788/RJC.2025.1819087>
18. Lisa Actis, Anand Srinivasan, Jose L. Lopez-Ribot, Anand K. Ramasubramanian and Joo L. Ong, *Journal of Materials Science: Materials in Medicine*, **26(215)**, 1(2015), <https://doi.org/10.1007/s10856-015-5538-8>
19. Angelo Frei, Anthony D. Verderosa, Alysha G. Elliott, Johannes Zuegg and Mark A. T. Blaskovich, *Nature Reviews Chemistry*, **7**, 202(2023), <https://doi.org/10.1038/s41570-023-00463-4>
20. Tikam Chand Dakal, Anu Kumar, Rita S. Majumdar, Vinod Yadav, *Frontiers in Microbiology*, **7**, 1831(2016), <https://doi.org/10.3389/fmicb.2016.01831>
21. Iris Xiaoxue Yin, Jing Zhang, Irene Shuping Zhao, May Lei Mei, Quanli Li, Chun Hung Chu, *International Journal of Nanomedicine*, **15**, 2555(2020), <https://doi.org/10.2147/IJN.S246764>
22. Sukumaran Prabhu and Eldho K. Poulose, *International Nano Letters*, **2(32)**, 1(2012), <https://doi.org/10.1186/2228-5326-2-32>

[RJC-9292/2025]